## Variability in Ranavirus Infection in Cell Lines of Cold-Blooded Vertebrates Gitanjali P. Talreja and Thomas B. Lentz Biotechnology Program, North Carolina State University, Raleigh, NC

## Abstract

Ranaviruses are large, double-stranded DNA viruses that infect cold-blooded vertebrates, such as amphibians, reptiles, and bony fish. The spread of these viruses has been Plaque implicated as a contributing factor in amphibian population declines in the United States, Assay Canada, and the United Kingdom. They are suspected factors in population declines of reptiles and fish and studies to evaluate this are ongoing. Understanding the susceptibility of different species to infection is an important step towards preventing viral spread, and therefore, reducing the impact of infection. The present study compares Ranavirus infection in cell lines derived from different host species. Cell lines from fish, Fathead We plated cells from We transferred a specific each cell line into Minnow and Epithelioma Papulosum Cyprini, amphibian, Xenopus Laevis, and reptile, volume of cells to a six wellseparate flasks. Terrapene Carolina and Vipera Russelli, were used. Cell lines were infected with Frog plate. Virus 3 strain because this is the most well-studied strain of Ranavirus. Multiple assays were used to determine the level of infectivity among the different host species. A plaque assay was performed with each cell line to determine if infection led to plaque formation, qPCR which is indicative of lytic replication. Periodic observation of infected cells monitored the Assav development of plaques in each cell line and was used to estimate time for plaque 1.95X10e6 celluine 1.3X10e5 celluine 5X10e2 celline. 99% formation. Additionally, viral load was measured by qPCR after infection of each cell type. Determining differences in viral load allows us to make inferences about alterations in the mechanisms of virus replication. Overall, these studies help us understand how the virus We plated 35,000 cells from each infects different species and the susceptibility of each to Ranavirus. We used the cell counter cell line into a 24-well plate. 4

itself.



https://en.wikipedia.org/wiki/Daboia https://en.wikipedia.org/wiki/Common\_box\_turtle https://en.wikipedia.org/wiki/Fathead\_minnow

to determine the concentration of cells.



cell lines were studied and each

cell line occupied 6 wells.

	Cell Lines	FHM
Days Post Infection	1	o-10 plaques
	3	10-40 plaques
	4	90-150 plaques
	6	90-150 plaques
	7	90-150 plaques
	9	Cells dead
	11	



All of the experimental data points fall in the range of the standard curve between 1X and (1\*10^-2) X.



## **qPCR Results**

 $R^2 = 0.9987$ 

**FHM** (+)

▲ EPC (+)

× A6 (+)

× VH (+)

• Cells (-)

There are a range of differences in the relative copy number of viral DNA between cell lines. The FHM cells contain the largest amount of viral y = -1.835ln(x) + 15.076 DNA, while VH cells contain the smallest amount of viral DNA. This 0.07 illustrates that the viral replication Standard Curve mechanisms differ between the cell lines. There was a very similar amount of viral DNA in FHM and EPC cells. There is greater replication of virus in FHM and EPC cells compared to the 0.02 —Log. (Standard Curve) other cell lines. This is evidence that 0.01 the Fathead Minnow species (FHM and EPC cells) is more susceptible to viral infection by the FV3 strain of **Ranavirus than Xenopus Laevis** (Amphibian) or Russell's Viper (Reptile).





is a 4-fold difference in relative copy number of viral DNA between the cell lines. In contrast, there is only a 2-fold difference between FHM and EPC cells.